

Assessing disease resistance in chickpeas through the bivariate analysis of Gaussian and binomial traits.

Aidan McGarty¹

Brian Cullis¹, Ahsan Asif² and Kristy Hobson²

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MMaED
Mixed Models and
Experimental Design Lab



UOW
AUSTRALIA

Mixed Models and Experiment Design Lab (MMaEDL)¹
National Institute for Applied Statistics Research Australia
University of Wollongong
amcgarty@uow.edu.au

Chickpea Breeding Australia²
NSW Department of Primary Industries | Agriculture

- **Motivation**
 - Context
 - Complexities
- **The statistics of the problem**
 - Choice of model
- **Results**
 - Parametric bootstrapping

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 - Field experiments are expensive, labour intensive and time consuming
- These factors lead to a complex statistical analysis

Motivation



Scenario

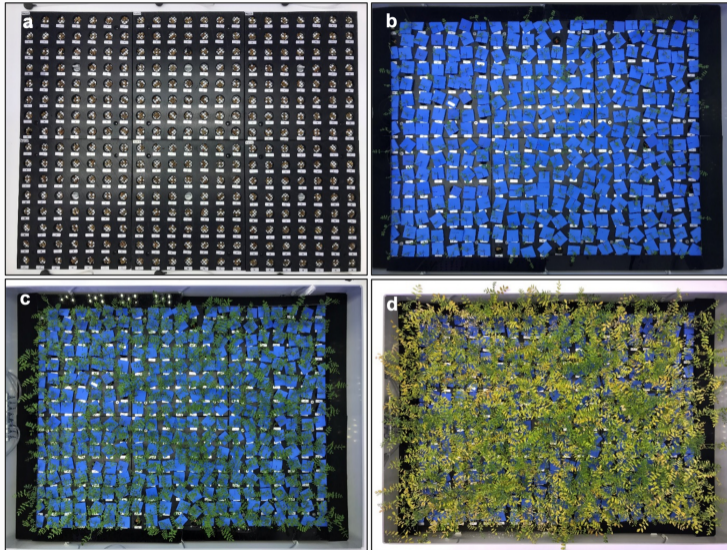
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- Hydroponics experiments provide high throughput low cost alternatives to field experiments

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Tank	Rack	HRange	HRow	Variety	TotalLeaf	DisLeaf
1	1	1	1	A	6	2
1	1	2	1	B	5	3
1	1	3	1	C		
1	1	4	1	D	11	6
1	1	5	1	F	13	9
1	1	6	1	A	15	7

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FCol	FRow	Variety	Risk1	Dead1	Risk2	Dead2	Risk3	Dead3	ExpectedLife
1	1	A	16	0	16	12	4	15	1.57
1	2	B	16	0	16	0	16	0	9.39
1	3	C	16	0	16	0	16	2	4.01
1	4	D	10	0	10	7	3	9	1.75
1	5	E	19	0	19	2	17	7	2.98
1	6	A	16	0	16	4	12	7	2.87

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 - Reflects the plot structure of each experiment
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 - Provides a reliable estimate of the genetic correlation between experiments
- We choose a bivariate generalised linear mixed model (GLMM) using ASReml-R

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- To overcome this the two data frames are merged by variety in a non-unique way

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WPlot	Variety	FCol	FRow	Tank	Rack	HRange	HRow	DL*	TL*	EL*
1	A	1	1	1	1	1	1	2	6	1.57
2	B	1	2	1	1	2	1	3	5	9.39
3	C	1	3	1	1	3	1			4.01
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*DL = Number of chlorotic leaves (hydroponic), *TL = Number of total leaves (hydroponic) and *EL = Expected lifetime (field)

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- WPlot arbitrarily joins field and hydroponic observations by common varieties, however as evident in WPlot 5 and 6 not all field and hydroponic observations can be matched

Model

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- We would also like to include random effects $\mathbf{u} = (\mathbf{u}_g^T, \mathbf{u}_{p_H}^T, \mathbf{u}_{p_F}^T)^T$ which are either shared genetic effects \mathbf{u}_g or non-shared peripheral effects \mathbf{u}_{p_H} and \mathbf{u}_{p_F} , for the hydroponic and field experiments respectively

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- We can then write the distributions $\mathbf{y}_H|\mathbf{u}$ and $\mathbf{y}_F|\mathbf{u}$ conditional on the random effects \mathbf{u} with associated probability density functions (PDF) $f_{Y_H|U}$ and $f_{Y_F|U}$

Model - Hydroponic Conditional Distribution

$$E(y_{H_i}|\mathbf{u}) = \mu_{H_i}$$

$$g(\mu_{H_i}) = \eta_{H_i} = \mathbf{x}_{H_i}^T \boldsymbol{\tau} + \mathbf{z}_{H_i}^T \mathbf{u}$$

$$V(y_{H_i}|\mathbf{u}) = \phi_H \mu_{H_i} \left(1 - \frac{\mu_{H_i}}{n_{TL_i}}\right)$$

- for $i = 1, \dots, n_H$ where $g(\cdot)$ is the logit link function
- \mathbf{x}_{H_i} and \mathbf{z}_{H_i} are indicator/covariate vectors of length c_X and c_Z relating to the fixed ($\boldsymbol{\tau}$) and random (\mathbf{u}) effects for the i th observation in the hydroponic experiment
- ϕ_H represents the dispersion parameter and n_{TL_i} are the binomial totals for the i th observation in the hydroponic experiment

Model - Field Conditional Distribution

$$E(y_{F_i}|\mathbf{u}) = \eta_{F_i} = \mathbf{x}_{F_i}^T \boldsymbol{\tau} + \mathbf{z}_{F_i}^T \mathbf{u}$$

$$V(y_{F_i}|\mathbf{u}) = \sigma_F^2$$

- for $i = 1, \dots, n_F$
- \mathbf{x}_{F_i} and \mathbf{z}_{F_i} are indicator/covariate vectors of length c_X and c_Z relating to the fixed ($\boldsymbol{\tau}$) and random (\mathbf{u}) effects for the i th observation in the field experiment
- σ_F^2 is the field residual variance

Random Effects

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$$\begin{bmatrix} \mathbf{u}_g \\ \mathbf{u}_{pH} \\ \mathbf{u}_{pF} \end{bmatrix} \sim N \left(\begin{bmatrix} \mathbf{0} \\ \mathbf{0} \\ \mathbf{0} \end{bmatrix}, \begin{bmatrix} \mathbf{G}_g & \mathbf{0} & \mathbf{0} \\ \mathbf{0} & \mathbf{G}_{pH} & \mathbf{0} \\ \mathbf{0} & \mathbf{0} & \mathbf{G}_{pH} \end{bmatrix} \right)$$

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- For convenience we allow $\boldsymbol{\kappa}$ to contain all parameters in \mathbf{G}_g , \mathbf{G}_{pH} and \mathbf{G}_{pF} along with ϕ_H and σ_F^2

Likelihood

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- The first term of which is not analytically tractable and hence requires an alternative to classical likelihood inference - therefore PQL is used to approximate the likelihood (Collins, 2008)

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- In the case of the non-genetic effects, we specify \mathbf{G}_{p_H} and \mathbf{G}_{p_F} as block diagonal matrices

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- This parameterisation is known as “reduced rank + diag” and is convenient as estimates of the λ_{ij} , $i = a, e$, $j = H, F$ cannot go outside the parameter space unlike the “ σ ” parameters in the unstructured parameterisation

Model - Recap

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- For the genetic effects, which are our main interest, we specify a variance structure which allows for covariance between experiments

```
asreml(fixed = cbind(DL,EL) ~ trait + trait:VarietyDrop,  
       random =~ rr(trait):vm(Variety, K) + at(trait,"EL"):vm(Variety, K) +  
               rr(trait):ide(Variety) + at(trait,"EL"):ide(Variety) +  
               at(trait, "DL"):Tank + at(trait, "DL"):Tank:Rack +  
               at(trait, "DL"):Tank:Rack:HRow + at(trait, "DL"):Tank:Rack:HRange +  
               at(trait, "EL"):FRow + at(trait, "EL"):FCol,  
       residual = ~id(WPlot):diag(trait),  
       data = df,  
       family = list(asr_binomial(total = "TL"), asr_gaussian()),  
       na.action = na.method(x="include",y="include"))
```

Results

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- To do so while also assessing potential bias in the PQL estimates, a known issue impacting parameter estimation (Breslow & Lin, 1995) we implement parametric bootstrapping

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- After 50 iterations simulations where $\Delta_{\text{norm}} > 0.001$ were excluded

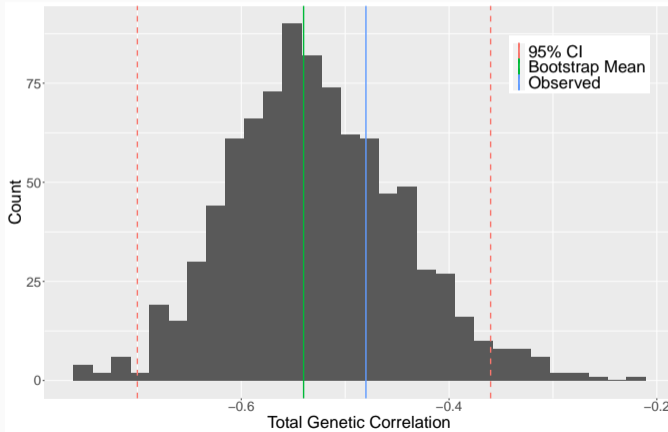
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Parametric Bootstrapping - Sampling Distribution

Parameter	Obs	BootMean
σ_{Tank}^2	0.082	0.077
σ_{Rack}^2	0.022	0.022
σ_{FRow}^2	0.000	0.014
σ_{FCol}^2	0.013	0.025
σ_{HRange}^2	0.038	0.035
σ_{HRow}^2	0.092	0.088
ψ_{a_F}	0.000	0.023
ψ_{e_F}	2.458	1.617
λ_{a_H}	0.151	0.335
λ_{a_F}	-1.074	-0.032
λ_{e_H}	0.376	0.997
λ_{e_F}	-0.591	-0.874
ϕ_H	1.095	0.965
σ_F^2	3.593	3.571

σ_j^2 , $j = \text{Tank, Rack, FRow, FCol, HRange and HRow}$ are the variance components associated with the peripheral random effects, ψ_{a_F} , ψ_{e_F} , λ_{a_H} , λ_{a_F} , λ_{e_H} and λ_{e_F} are the genetic parameters and ϕ_H and σ_F^2 are the hydroponic dispersion and field residual variance parameter respectively

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- The resulting analysis gave a quantification of the level of agreement between PRR resistance for the two experiments
- Parametric bootstrapping provided assessment of parameter estimation bias and indicated the associated 95% bootstrap CI for the total genetic correlation does not contain 0

Rapid and high throughput hydroponics phenotyping method for evaluating chickpea resistance to *Phytophthora* root rot


Muhammad A. Asif^{1*}, Sean L. Bithell², Ramethaa Pirathiban³, Brian R. Cullis³, David Hughes³, Aidan McGarty³, Nicole Dron² and Kristy Hobson¹

¹Chickpea Breeding Australia, New South Wales Department of Primary Industries, Tamworth Agricultural Institute, Tamworth, New South Wales, Australia

²New South Wales Department of Primary Industries, Tamworth Agricultural Institute, Tamworth, New South Wales, Australia

³Centre for Biometrics and Data Science for Sustainable Primary Industries, National Institute for Applied Statistics and Research Australia, School of Mathematics and Applied Statistics, University of Wollongong, Australia ›

*Corresponding author: ahsan.asif@dpi.nsw.gov.au




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Mapping resistance to phytophthora root rot identifies independent loci from cultivated (*cicer arietinum* L.) and wild (*cicer echinospermum* p.h. davis) chickpea.

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