Estimation of the amplicon methylation pattern distribution from bisulphite sequencing data

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Cytosines in a genome, particularly those in the combination CpG, can undergo an epigenetic change called *methylation* 



Methylation patterns

- can play a role in cell development
- can play a role in determining phenotype
- can be a response to environmental factors
- can change from cell-type to cell-type in an organism

A population of cells of a given type in a given organism defines a probability distribution over methylation patterns

e.g. for 3 nearby CpG sites

k	pattern*	$\operatorname{Prob}(K=k) = \theta_k$	
			1.0 -
1	000	0.72	
2	001	0.12	0.8 -
3	010	0.00	0.6 -
4	011	0.00	0.4 -
5	100	0.10	0.2 -
6	101	0.00	
7	110	0.01	
8	111	0.05	1 2 3 4 5 6 7 8
			K

\*0 = unmethyated 1 = methylated

# Methylation patterns are measured directly in the laboratory via Bisulphite Conversion







We have developed the R Bioconductor Package **MPFE**:

- Methylation Patterns Frequency Estimation
- Inputs (*n* cytosines)
- A table of methylation pattern counts  $y_k$ , where  $1 \le k \le 2^n$  labels the  $2^n$  different methylation patterns
- The non-conversion rate  $\varepsilon$
- The sequencing error rate  $\eta$ , either global or site-dependent



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- The non-conversion rate  $\varepsilon$
- The sequencing error rate  $\eta$ , either global or site-dependent
- Outputs
- A table of patterns and their estimated frequencies  $\theta_k$
- List of spurious patterns called
- Plots comparing the observed and estimated frequencies



## Synthetic Data



sequencing error rate  $\eta = 0.005$ 25 patterns, 9 are called spurious



### How does it work?

Given:

- *n* CpG sites,  $k = 1, 2, ..., 2^n$  possible patterns
- true distribution over patterns to be estimated

$$\Pr(K = k) = \theta_k$$

• non-conversion rate  $\varepsilon$  and read error rate  $\eta$ 

The probability of a true pattern k registering as pattern l is

$$\Pr(L=l|K=k) = M_{kl}$$

where the  $2^{n} \times 2^{n}$  matrix *M* is (after a little bit of algebra)

$$M = E \otimes E \otimes \ldots \otimes E, \qquad E = \begin{pmatrix} 1 - \varepsilon - \eta + 2\varepsilon\eta & \varepsilon + \eta - 2\varepsilon\eta \\ \eta & 1 - \eta \end{pmatrix}$$
*n* times

Then the probability a read registers as pattern number *l* is

$$\Pr(L=l) = \sum_{k=1}^{2^{n}} \Pr(K=k) \Pr(L=l | K=k) = \sum_{k=1}^{2^{n}} \theta_{k} M_{kl}$$

The distribution of read counts  $Y_1$ ,  $Y_2$ , ...,  $Y_{2^n}$  for patterns  $l = 1, ..., 2^n$  out of a total of N reads is a multinomial distribution:

$$\Pr(\mathbf{Y} = \mathbf{y}|\theta) = \frac{N!}{y_1! y_2! \dots y_{2^n}!} \prod_{l=1}^{2^n} \left(\sum_{k=1}^{2^n} \theta_k M_{kl}\right)^{y_l}$$

... which enables a maximum likelihood estimate of the underlying distribution  $\theta$  from

$$L(\theta | \mathbf{Y} = \mathbf{y}) = \log\left(\Pr\left(\mathbf{Y} = \mathbf{y} | \theta\right)\right) \propto \sum_{l=1}^{2^{n}} y_{k} \log\left(\sum_{k=1}^{2^{n}} \theta_{k} M_{kl}\right)$$

subject to the important constraints

$$\sum_{k=1}^{2^n} \theta_k = 1, \quad \theta_k \ge 0.$$

- Implemented in R using the function constrOptim()
- For realistic data the estimate of  $\theta_k$  is generally on the boundary of the constraint  $\theta_k \ge 0$ . I.e. there are many 'observed' patterns which turn out to be spurious.

## Synthetic Data



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## Synthetic Data



total number of reads = 2000 non-conversion rate  $\varepsilon$  = 0.008 sequencing error rate  $\eta$  = 0.005 25 patterns, 9 are called spurious Lin et al. BMC Bioinformatics (2015) 16:145 DOI 10.1186/s12859-015-0600-6

#### BMC Bioinformatics

#### METHODOLOGY ARTICLE

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# Estimation of the methylation pattern distribution from deep sequencing data

Peijie Lin<sup>1</sup>, Sylvain Forêt<sup>2</sup>, Susan R Wilson<sup>1,3</sup> and Conrad J Burden<sup>1</sup>

#### Abstract

**Background:** Bisulphite sequencing enables the detection of cytosine methylation. The sequence of the methylation states of cytosines on any given read forms a methylation pattern that carries substantially more information than merely studying the average methylation level at individual positions. In order to understand better the complexity of DNA methylation landscapes in biological samples, it is important to study the diversity of these methylation patterns. However, the accurate quantification of methylation patterns is subject to sequencing errors and spurious signals due to incomplete bisulphite conversion of cytosines.

**Results:** A statistical model is developed which accounts for the distribution of DNA methylation patterns at any given locus. The model incorporates the effects of sequencing errors and spurious reads, and enables estimation of the true underlying distribution of methylation patterns.

**Conclusions:** Calculation of the estimated distribution over methylation patterns is implemented in the R Bioconductor package MPFE. Source code and documentation of the package are also available for download at http://bioconductor.org/packages/3.0/bioc/html/MPFE.html.

Keywords: DNA methylation, Bisulfite sequencing, DNA methylation patterns, Epiallele

#### Background

Epigenetic regulations are involved in a broad range of biological processes, including development, tissue home-ostasis, learning and memory, as well as various diseases such as obesity and cancer [1-3].

DNA methylation is one of the best studied epigenetic molecular mechanisms. It consists of the addition of a methyl group to the cytosine residues (C) of a DNA molecule. In animals, DNA methylation usually takes place in the CpG context: cytosines followed by a guanine (G) residue.

DNA methylation modulates gene expression through

The diverse and subtle effects of DNA methylation enable a given genome to produce different phenotypic outputs as part of a developmental program or in response to environmental factors. This has fundamental implications at the organismal level, where DNA methylation plays an important role in phenotypic plasticity [6]. This is also important at the cellular level to create diverse cell types, tissues and organs all based on the same genome. DNA methylation patterns can thus change from one cell type to another or within a cell under different conditions [7].

The diversity of methylation patterns in a sample can be



### distribution from deep sequencing data

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#### Abstract

Background: Bis methylation states information than in the complexity of methylation patte and spurious signa Results: A statisti given locus. The in the true underlyin Conclusions: Cal Bioconductor pac http://bioconduct Keywords: DNA

#### Background

Epigenetic regulati biological processes ostasis, learning an such as obesity and DNA methylation molecular mechan methyl group to t molecule. In anim place in the CpG co (G) residue. DNA methylation SCIENTIFIC REPORTS



OPEN EGFR gene methylation is not involved in Royalactin controlled phenotypic polymorphism in honey bees

R. Kucharski, S. Foret & R. Maleszka

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Sylvain Forêt RSB, ANU Paul Lin UNSW Susan Wilson ANU & UNSW

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